## Example timeline of a herd closure and medication program for Mycoplasma hyopneumoniae eradication using aerosolization exposure with concurrent off-site M. hyopneumoniae negative breeding project

Modified from Yeske P, Betlach A, Hoist S, Pieters M. Eradication of *Mycoplasma hyopneumoniae* from pig herds. In: Maes D, Sibila M, Pieters M. eds. Mycoplasmas in swine. 1st ed. Acco Nederland, The Netherlands 2020; 229–245. (Table 12.2 (p. 239)) and McDowell E, Pieters M, Spronk T, Nerem J, Fano E, Dee S, Sponheim A. Duration of *Mycoplasma hyopneumoniae* detection in pigs following purposeful aerosol exposure. Vet Microbiol 2023; 282:109758. (Table 1).

Compiled by Sponheim, Fano, and Pieters, 2023.

Microsoft Word file available upon request at <a href="mailto:ahmhproadmap.us@boehringer-ingelheim.com">ahmhproadmap.us@boehringer-ingelheim.com</a>.

HERD CLOSURE	EVENT
-35	Deep tracheal secretion sampling to identify donors for exposure
-31	Treatment of potential donor animals with antibiotic of known <i>M. hyopneumoniae</i> resistance ( <i>i.e.</i>
J1	compounds with activity directed at cell wall)
-29	Naïve M. hyopneumoniae replacement gilts received at GDU considered part of "herd" during
	closure
-28	Lung homogenate preparation and exposure at sow farm, collect bronchial swab(s) and obtain <i>M. hyopneumoniae</i> sequence for potential future comparison
-21	Lung homogenate exposure at GDU(s) considered part of "herd" during closure
-21	Concurrent off-site gilt breeding project: Begin receiving naïve <i>M. hyopneumoniae</i> replacement gilts at GDU not considered part of "herd" during closure (assumes receiving gilts around 3 weeks of age). Maintain replacement gilt vaccination program.
0	Deep tracheal secretion sampling from replacement gilt populations to confirm effective exposure
	(may be as early as 14 days post-exposure)
	*Sow population sample type will depend on historical exposure status. Historical acclimation
	program: serum and ELISA with interpretation of vaccine program; Naïve: deep tracheal secretion and PCR
0	Begin herd closure, including <i>M. hyopneumoniae</i> exposed GDU(s). There must be NO
	additions to resident/gilt pool populations, including saving back internal replacement gilts, or
	bringing in teaser boars. Limit sow herd culling to maintain breed targets. Maintain piglet and
	replacement gilt vaccination program.
0	Whole herd vaccination
30	Whole herd vaccination
60	Whole herd vaccination
168	Concurrent off-site gilt breeding project: Begin breeding M. hyopneumoniae negative gilts (30
100	week of age)
182	Wash and disinfect gestation and on-site GDU barns***
189	Whole sow herd medication***
189	Medication of all piglets on site***
189	Piglet treatment at birth***
203	Begin new whole herd antibiotic***
203	Begin piglet treatment at 14 days of age (treat all piglets 14 days of age and older)***
217	Last piglets treated at birth***
231	Last piglet treatment at 14 days of age***
240	Deep tracheal secretion sampling of breeding herd population to confirm eradication program success
240	<b>End herd closure.</b> If diagnostic criteria met, confirm program success and herd opens. If not,
268	follow contingency plans (ie: extend closure, resample, etc).  M. hyopneumoniae confirmed negative bred replacement gilts enter breeding herd (assumes begi
200	entering at ~100 days gestation). Allows additional time before farrowing if needed for contingency plan.
>268 ** Activities	Monitoring clinical signs and targeted testing to continue confirming project success in <i>M. hyopneumoniae</i> naive, unvaccinated replacement gilts. Piglet and replacement gilt vaccination commonly maintained, even after a successful eradication program. specific to concurrent off-site gilt breeding project shaded dark grey
	with herd veterinarian and/or manufacturers

## References:

- 1. Holst S., Yeske, P., and Pieters, M. (2015). Elimination of *Mycoplasma hyopneumoniae* from breed-to-wean farms: A review of current protocols with emphasis on herd closure and medication. Journal of Swine Health and Production, 23(6), 321-330.
- 2. Yeske, P. (2007). Mycoplasma elimination strategies. Proceedings American Association of Swine Veterinarians, Orlando, Florida, USA, 367-370.
- 3. Alfonso, A., Geiger, J.O., Freixes, C., Fonz, J.C., and Torremorell, M. (2004). *Mycoplasma hyopneumoniae* and PRRSV elimination in a 1700 sow multi-site system. Proceedings Allen D. Leman Swine Conference, St. Paul, MN, USA, 13.
- 4. Robbins, R., Betlach, A., Mondragon-Evans, M., and Pieters, M. (2019). Development of a herd-specific lung homogenate for exposure to *Mycoplasma hyopneumoniae* under field conditions. Journal of Swine Health and Production, 27(4), 221-227.
- 5. Figueras Gourgues, S., Fano, E., Alegre Sabate, A., Lopez Grasa, E., Hernandez Caravaca, I., Garcia Vazquez, F. A., Rodriquez Vega, V., and Garcia-Morante, B. (2020). Assessment of nebulization technology for gilt exposure to *Mycoplasma hyopneumoniae* as an acclimation strategy. Journal of Swine Health and Production, 28(6), 294-301.
- 6. McDowell E, Pieters M, Spronk T, Nerem J, Fano E, Dee S, Sponheim A. Duration of *Mycoplasma hyopneumoniae* detection in pigs following purposeful aerosol exposure. Vet Microbiol 2023;282:109758. (Table 1)
- 7. Sponheim, A., Alvarez, J., Fano, E., Rovira, A., McDowell, E., Toohill, E., Dalquist, L., Pieters, M. A diagnostic approach to confirm *Mycoplasma hyopneumoniae* "Day zero" for pathogen eradication. Prev Vet Med 2023;221:106057.
- 8. Sponheim, A., Munoz-Zanzi, C., Fano, E., Polson, D., and Pieters, M. (2021). Pooled-sample testing for detection of *Mycoplasma hyopneumoniae* during late experimental infection as a diagnostic tool for a herd eradication program. Preventive Veterinary Medicine, 189, 105313.
- 9. Yeske P, Betlach A, Hoist S, Pieters M. Eradication of *Mycoplasma hyopneumoniae* from pig herds. In: Maes D, Sibila M, Pieters M. eds. Mycoplasmas in swine. 1st ed. Acco Nederland, The Netherlands 2020; 229–245. (Table 12.2 (p. 239))